

SUPPLEMENT TO:

Clinical Pharmacogenetics Implementation Consortium (CPIC) Guideline for *HLA* Genotype and Use of Carbamazepine and Oxcarbazepine: 2017 Update

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GUIDELINE UPDATES

The Clinical Pharmacogenetics Implementation Consortium (CPIC) guideline for *HLA* genotype and use of carbamazepine and oxcarbazepine is published in full on the CPIC website (<http://cpicpgx.org>) and the PharmGKB website (www.pharmgkb.org). Information will be reviewed periodically and updated guidelines published online.

UPDATES IN SUPPLEMENT

- Updated literature search from January 2013 to June 2016 for *HLA-B*15:02* and carbamazepine.
- Expanded literature search to include *HLA-A*31:01* and oxcarbazepine.
- Updated evidence linking *HLA-B*15:02* to carbamazepine-induced Stevens-Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN).
- Added evidence linking *HLA-B*15:02* to oxcarbazepine-induced SJS/TEN and evidence linking *HLA-A*31:01* to carbamazepine-induced SJS/TEN, drug reaction with eosinophilia and systemic symptoms (DRESS), and maculopapular exanthema (MPE).
- Added a section on the proposed mechanism for carbamazepine-induced SJS/TEN in *HLA-B*15:02* positive patients.
- Added resources to facilitate incorporation of *HLA* genotype results into electronic health records with clinical decision support and updated allele frequency information (see <https://cpicpgx.org/guidelines/guideline-for-carbamazepine-and-hla-b/>).

LITERATURE REVIEW

A search was conducted on the PubMed database (1966 to June 17, 2016) for keywords ([HLA] AND [carbamazepine]), ([HLA] AND [oxcarbazepine]), ([HLA] AND [eslicarbazepine]), and ([HLA] AND [lamotrigine]). The ([HLA] AND [carbamazepine]) search yielded 238 articles, the ([HLA] AND [oxcarbazepine]) search yielded 14 articles, the ([HLA] AND [eslicarbazepine]) search yielded two articles, and the ([HLA] AND [lamotrigine]) search yielded 30 articles. Study inclusion criteria included publications that explored the association between *HLA-B*15:02* or *HLA-A*31:01* genotypes and severe cutaneous adverse drug reactions

with any of the aforementioned antiepileptics. Non-English manuscripts were excluded. Following application of these inclusion criteria, 82 publications were reviewed.

A table of frequencies of the *HLA-B*15:02* and *HLA-A*31:01* alleles in different ethnic populations around the world was assembled from several sources (see ***HLA-A and HLA-B Allele Frequency Table*** online). Frequencies were included from the Allele Frequencies in Worldwide Populations website (<http://www.allelefrequencies.net/>), which lists frequency data for *HLA* alleles from over 200 different samples and populations. Allele frequencies were also obtained by conducting a search of the PubMed database (2000 to 2016) using the following criteria: ([*HLA-B*1502*] AND [frequency]) and ([*HLA-A*3101*] AND [frequency]). Studies from both sources were considered for inclusion if the following criteria were met: 1) the ethnicity of the population was clearly indicated; 2) allele frequencies were reported; and 3) the sample population consisted of at least 100 individuals.

OTHER CONSIDERATIONS

Allele Frequency vs. Allele Carriage Rate

Representation of *HLA* in a given population can be described in terms of either allele frequency (the total number of copies of the allele in the relevant population), or by allele carriage rate (the percentage of individuals who have the allele in the population or prevalence). This concept differs from other genes because *HLA* is inherited in a co-dominant fashion and to take into account those who are homozygous or have two copies of a given *HLA* allele. The representation of homozygosity in any given population may have been driven by a number of evolutionary factors that select against this (“the heterozygous advantage”) (1, 2). For carbamazepine-induced SJS/TEN and abacavir hypersensitivity, there is no current evidence to suggest a gene-dose effect or that carrying more than one copy of the *HLA* risk allele is associated with a higher risk; however, for some other phenotypes (e.g., dapsone hypersensitivity and *HLA-B*13:01*), being homozygous portends a higher risk of disease (3).

Proposed Mechanism for HLA-B*15:02-mediated SJS/TEN in Response to Carbamazepine

Current research suggests that carbamazepine binds non-covalently to the B pocket of the HLA-B*15:02 peptide binding groove and that the B pocket residues Arg62, Asn63, Ile95, and Leu156 contribute to drug-HLA interactions (4). Other members of the B75 serotype implicated in carbamazepine SJS/TEN (as described in the following section) also share these B pocket residues. However, other mechanisms by which carbamazepine and its metabolites bind to the HLA molecules cannot be discounted and may account for differences in the presentation of clinical symptoms between different patients (5).

T-cell receptor (TCR) sequencing of blister-fluid derived T cells from patients with *HLA-B*15:02*-associated carbamazepine SJS/TEN has identified a shared CD8⁺ TCR clonotype that bears a common CDR3 sequence that is found in the peripheral blood of carbamazepine SJS/TEN patients but not in peripheral blood of drug tolerant controls or in blister fluid from patients with SJS/TEN secondary to another drug. Thus, although the crystal structure of HLA-B*15:02 bound to peptide drug and TCR has not been solved and the role of a peptide remains to be determined, the immunopathogenesis of *HLA-B*15:02*-associated carbamazepine SJS/TEN likely depends upon the concomitant involvement of both a specific HLA allotype and a specific TCR clonotype (6).

HLA-B75 Serotype

Relevant to the proposed mechanism for *HLA-B*15:02*-associated carbamazepine SJS/TEN, HLA molecules of the same B75 serotype (e.g., HLA-B*15:08, HLA-B*15:11 and HLA-B*15:21) with similar peptide binding properties have also been associated with carbamazepine SJS/TEN, particularly HLA-B*15:11 in populations such as Japanese and Koreans where HLA-B*15:02 is less prevalent (7-9). Currently, carbamazepine-induced SJS/TEN has not been associated with less frequently carried B75 serotype alleles such as *HLA-B*15:30* and *HLA-B*15:31*; however, given the structural similarity and shared peptide binding properties, the risk for this should also be considered. If a patient developed SJS/TEN despite a negative *HLA-B*15:02* and/or *HLA-A*31:01* result, full *HLA-B* typing may provide further insight, particularly if the test reveals the presence of another B75 serotype allele.

Other Aromatic Anticonvulsants

Several drugs structurally similar to carbamazepine and oxcarbazepine have also been associated with drug-induced cutaneous adverse reactions in *HLA-B*15:02* positive patients or are thought to be associated with greater risk, including phenytoin, eslicarbazepine acetate, and lamotrigine. For a detailed discussion of *HLA-B*15:02* and phenytoin, please refer to the previously published CPIC guideline (10). Eslicarbazepine acetate is an antiepileptic drug used in Europe and America. It is a prodrug which is activated to (S)-licarbazepine, an active metabolite of oxcarbazepine. As of this report, there have been no cases reported of eslicarbazepine-induced cutaneous adverse reactions associated with *HLA-B*15:02*; however, based on its structural similarity to oxcarbazepine, caution should be used in patients positive for *HLA-B*15:02*. Lamotrigine has also been associated with SJS/TEN, particularly with rapid dose escalation or when used in combination with valproic acid. A 2015 meta-analysis involving four studies in Han Chinese patients found a statistical association between *HLA-B*15:02* and lamotrigine-induced SJS/TEN, with an odds ratio of 4.98 (11).

Available Genetic Test Options and Interpretation

Commercially available genetic testing options change over time. Information that may assist in evaluating options is available below. Furthermore, the Genetic Testing Registry (GTR) provides a central location for voluntary submission of genetic test information by providers and is available at <http://www.ncbi.nlm.nih.gov/gtr/>.

HLA alleles are extremely diverse and typically consist of numerous nucleotide and resultant amino acid substitutions. Comparison of nucleotide sequences for a reference *HLA-B* allele with that of *HLA-B*15:02* reveals 42 differences within the open reading frame of the gene (**Supplemental Figure S1**). These nucleotide sequence differences translate to a peptide exhibiting 27 amino acid substitutions in the variant allele (**Supplemental Figure S2**). Comparison of the *HLA-A*31:01* allele with the reference *HLA-A*01:01* reveals 46 differences within the open reading frame of the gene (**Supplemental Figure S3**). These nucleotide sequence differences translate to a peptide exhibiting 33 amino acid substitutions in the variant allele (**Supplemental Figure S4**).

A variety of companies provide clinical testing services for the detecting of *HLA-B*15:02* and *HLA-A*31:01*. They primarily employ two different detection methods. One is direct sequencing of the gene. Alleles are assigned by comparison of the sequence to the known variants that define *HLA-B*15:02* or *HLA-A*31:01* and reported as the diplotype of *HLA-B* or *HLA-A* alleles, respectively.

Genotyping is another common approach in which the sequence variants that define *HLA-B*15:02* and *HLA-A*31:01* are directly detected through a panel of DNA tests. Allele specific polymerase chain reaction (PCR) is commonly employed where PCR primers specific for each nucleotide variant are used. The PCR products can then be detected using gel electrophoresis or other methods. A variety of other genotyping methods may also be used to directly detect each of the nucleotide variants for *HLA-B*15:02* and *HLA-A*31:01*. As the test is specific for *HLA-B*15:02* or *HLA-A*31:01*, the test will only report its presence or absence as opposed to the full diplotype available through sequencing.

Another option is the genotyping of one or more single nucleotide polymorphisms (SNPs) that are near the *HLA-B* locus and in linkage disequilibrium with the *HLA-B*15:02* allele. However, as this test is indirect and depends upon linkage disequilibrium which may vary between different populations, it may have lower accuracy. It also requires genotyping and may not be any faster or less expensive than genotyping of the specific defining variants.

LEVELS OF EVIDENCE

The evidence summarized in **Supplemental Tables S1 and S2** is graded on a scale of high, moderate, and weak, based upon level of evidence:

High: Evidence includes consistent results from well-designed, well-conducted studies.

Moderate: Evidence is sufficient to determine effects, but the strength of the evidence is limited by the number, quality, or consistency of the individual studies, generalizability to routine practice, or indirect nature of the evidence.

Weak: Evidence is insufficient to assess the effects on health outcomes because of limited number or power of studies, important flaws in their design or conduct, gaps in the chain of evidence, or lack of information.

Based on the levels of evidence for major findings, the strength of therapeutic recommendations are assigned accordingly (**Tables 2 and 3, main manuscript**).

STRENGTH OF RECOMMENDATIONS

CPIC uses a slight modification of a transparent and simple system for recommendations adopted from the rating scale for evidence-based recommendations on the use of antiretroviral agents (12):

Strong: The evidence is high quality and the desirable effects clearly outweigh the undesirable effects.

Moderate: There is a close or uncertain balance as to whether the evidence is high quality and the desirable effects clearly outweigh the undesirable effects.

Optional: The desirable effects are closely balanced with undesirable effects, or the evidence is weak or based on extrapolations. There is room for differences in opinion as to the need for the recommended course of action.

No recommendation: There is insufficient evidence, confidence, or agreement to provide a recommendation to guide clinical practice at this time.

RESOURCES TO INCORPORATE PHARMACOGENETICS INTO AN ELECTRONIC HEALTH RECORD WITH CLINICAL DECISION SUPPORT

Clinical decision support (CDS) tools integrated into electronic health records (EHRs) can help guide the optimal use of pharmacogenetic test results at the point of care (13-17). Please refer to the CPIC website for this guideline (<https://cpicpgx.org/guidelines/guideline-for-carbamazepine-and-hla-b/>) for resources to support the adoption of this guideline's recommendations into an EHR. Based on the capabilities of various EHRs and local preferences, we recognize that approaches may vary across organizations. Our intent is to synthesize foundational knowledge that provides a common starting point for incorporating the use of *HLA* genotype results to guide the rational use of carbamazepine and oxcarbazepine in an EHR.

Effectively incorporating pharmacogenetic information into an EHR to optimize drug therapy should have some key attributes. Pharmacogenetic results, an interpreted phenotype, and a concise interpretation or summary of the result must be documented in the EHR (18, 19). To incorporate a phenotype in the EHR in a standardized manner, genotype test results provided by the laboratory must be consistently translated into an interpreted phenotype (**Table 1, main manuscript**). Because clinicians must be able to easily find the information, the interpreted phenotype may be documented as a problem list entry, in a patient summary section, or in the adverse drug reaction section; these phenotypes are best stored in the EHR at the “person level” that links to both the inpatient and outpatient record rather than at the date-centric “encounter level.” Additionally, results should be entered as standardized and discrete terms to facilitate using them to provide point-of-care CDS (13, 20).

Because pharmacogenetic results have lifetime implications of clinical significance that may expand as more knowledge becomes available, results should be placed into a section of the EHR that is accessible independent of the test result date to allow clinicians to quickly find the result at any time after it is initially placed in the EHR. To facilitate this process, CPIC provides gene-specific figures and tables that illustrate how *HLA* pharmacogenetic test results could be entered into an EHR, example EHR consultation/genetic test interpretation language, and widely used nomenclature systems for genes and drugs relevant to the guideline

(see the following online resources: ***HLA-B* Genotype Table**, ***HLA-A* Genotype Table**, ***HLA* Gene Resource Mappings Table**, **Carbamazepine Drug Resource Mappings Table**, and **Oxcarbazepine Drug Resource Mappings Table**).

Point-of-care CDS should be designed to effectively notify clinicians of prescribing implications at any time after the test result is entered into the EHR. CPIC also provides gene-drug specific tables and example pre- and post-alert language that provide guidance to achieve these objectives with diagrams that illustrate how point-of-care CDS should be entered into the EHR (see the following online resources: **Carbamazepine Pre- and Post-test Alerts and Flow Charts** and **Oxcarbazepine Pre-and Post-test Alerts and Flow Charts**).

TABLE S1. EVIDENCE LINKING *HLA-B*15:02* GENOTYPE WITH CARBAMAZEPINE- AND OXCARBAZEPINE-INDUCED CUTANEOUS ADVERSE REACTIONS

Type of Experimental Model	Clinical Phenotype	Major Findings	References	Level of Evidence ^a
Carbamazepine and <i>HLA-B*15:02</i>				
In vitro	N/A	PBMCs from carbamazepine-induced SJS/TEN patients, all <i>HLA-B*15:02</i> positive, have significantly higher levels of interferon gamma and granulysin when cultured with carbamazepine, compared to carbamazepine-tolerant patients (2 <i>HLA-B*15:02</i> positive and 9 <i>HLA-B*15:02</i> negative).	Ko, <i>et al.</i> 2011 (21)	Moderate
In vitro	SJS/TEN	Patients with carbamazepine-induced SJS/TEN and <i>HLA-B*15:02</i> positive mounted a cytotoxic T lymphocyte response. This response was absent in carbamazepine-tolerant <i>HLA-B*15:02</i> positive patients.	Wei, <i>et al.</i> 2012 (4)	Moderate
Clinical	SJS/TEN	Prospective screening of <i>HLA-B*15:02</i> reduces the incidence of carbamazepine-induced SJS/TEN compared to historical data.	Chen, <i>et al.</i> 2011 (22) Chen, <i>et al.</i> 2014 (23)	High
Clinical	SJS/TEN	Significant association between <i>HLA-B*15:02</i> genotype and patients with carbamazepine-induced SJS/TEN compared to carbamazepine-tolerant patients and/or healthy controls.	Supports statement: Chung, <i>et al.</i> 2004 (24) Hung, <i>et al.</i> 2006 (25) Man, <i>et al.</i> 2007 (26)	High

			<p>Locharernkul, <i>et al.</i> 2008 (27)</p> <p>Mehta, <i>et al.</i> 2009 (28)</p> <p>Tassaneeyakul, <i>et al.</i> 2010 (7)</p> <p>Wu, <i>et al.</i> 2010 (29)</p> <p>Chang, <i>et al.</i> 2011 (30)</p> <p>Then, <i>et al.</i> 2011 (31)</p> <p>Wang, <i>et al.</i> 2011 (32)</p> <p>Zhang, <i>et al.</i> 2011 (33)</p> <p>Kulkantrakorn, <i>et al.</i> 2012 (34)</p> <p>Shi, <i>et al.</i> 2012 (35)</p> <p>Neuman, <i>et al.</i> 2012 (36)</p> <p>Amstutz, <i>et al.</i> 2013 (37)</p> <p>He, <i>et al.</i> 2013 (38)</p> <p>Cheung, <i>et al.</i> 2013 (39)</p> <p>Lin, <i>et al.</i> 2013 (40)</p>	
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			<p>Aggarwal, <i>et al.</i> 2014 (41)</p> <p>Chong, <i>et al.</i> 2014 (42)</p> <p>Khor, <i>et al.</i> 2014 (43)</p> <p>Kwan, <i>et al.</i> 2014 (44)</p> <p>Sun, <i>et al.</i> 2014 (45)</p> <p>Genin, <i>et al.</i> 2014 (46)</p> <p>Hsiao, <i>et al.</i> 2014 (47)</p> <p>Toh, <i>et al.</i> 2014 (48)</p> <p>Wang, <i>et al.</i> 2014 (49)</p> <p>Nguyen, <i>et al.</i> 2015 (50)</p> <p>Yang, <i>et al.</i> 2015 (51)</p> <p>Teh, <i>et al.</i> 2016 (52)</p> <p>Indeterminate (inadequate statistical power to detect low frequency variant):</p> <p>Alfirevic, <i>et al.</i> 2006 (53)</p>	
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			<p>Kashiwagi, <i>et al.</i> 2008 (54)</p> <p>Kaniwa, <i>et al.</i> 2008 (55)</p> <p>Kano, <i>et al.</i> 2008 (56)</p> <p>Ikeda, <i>et al.</i> 2010 (57)</p> <p>Kaniwa, <i>et al.</i> 2010 (8)</p> <p>Kim, <i>et al.</i> 2011 (9)</p> <p>Niihara, <i>et al.</i> 2012 (58)</p> <p>Park, <i>et al.</i> 2016 (59)</p>	
Clinical	DRESS/MPE	No significant association between <i>HLA-B*15:02</i> genotype and patients with carbamazepine-induced non-SJS/TEN cutaneous adverse drug reaction compared to carbamazepine-tolerant patients and/or healthy controls.	<p>Alfirevic, <i>et al.</i> 2006 (53)</p> <p>Hung, <i>et al.</i> 2006 (25)</p> <p>Man, <i>et al.</i> 2007 (26)</p> <p>Kashiwagi, <i>et al.</i> 2008 (54)</p> <p>Kano, <i>et al.</i> 2008 (56)</p> <p>Ikeda, <i>et al.</i> 2010 (57)</p> <p>Wu, <i>et al.</i> 2010 (29)</p> <p>Wang, <i>et al.</i> 2011 (32)</p> <p>Niihara, <i>et al.</i> 2012 (58)</p>	High

			<p>Amstutz, <i>et al.</i> 2013 (37)</p> <p>Li, <i>et al.</i> 2013 (60)</p> <p>Lin, <i>et al.</i> 2013 (40)</p> <p>Sun, <i>et al.</i> 2014 (45)</p> <p>Hsiao, <i>et al.</i> 2014 (47)</p> <p>Locharernkul, <i>et al.</i> 2008 (27)</p> <p>Chong, <i>et al.</i> 2014 (42)</p> <p>Nguyen, <i>et al.</i> 2015 (50)</p>	
Clinical	SJS/TEN	Cases of patients with carbamazepine-induced SJS/TEN and <i>HLA-B*15:02</i> genotype.	<p>Lonjou, <i>et al.</i> 2006 (61)</p> <p>Odueyungbo, <i>et al.</i> 2010 (62)</p> <p>Elzagallaai, <i>et al.</i> 2011 (63)</p> <p>Wang, <i>et al.</i> 2012 (64)</p> <p>Techasatian, <i>et al.</i> 2015 (65)</p> <p>Tan, <i>et al.</i> 2015 (66)</p> <p>Bellon, <i>et al.</i> 2016 (67)</p>	Moderate

Oxcarbazepine and <i>HLA-B*15:02</i>				
Clinical	SJS/TEN	Significant association between <i>HLA-B*15:02</i> genotype and patients with oxcarbazepine-induced SJS/TEN compared to oxcarbazepine-tolerant patients or healthy controls.	<p>Supports statement: Chen, <i>et al.</i> 2017 (68)</p> <p>Indeterminate (inadequate statistical power to detect low frequency variant): Amstutz, <i>et al.</i> 2013 (37)</p>	High
Clinical	DRESS/MPE	No significant association between <i>HLA-B*15:02</i> genotype and patients with oxcarbazepine-induced non-SJS/TEN cutaneous adverse drug reaction compared to oxcarbazepine-tolerant patients and/or healthy controls.	<p>Supports statement: Hu, <i>et al.</i> 2011 (69) He, <i>et al.</i> 2012 (70) Lv, <i>et al.</i> 2013 (71) Sun, <i>et al.</i> 2014 (45) Wang, <i>et al.</i> 2014 (72) Chen, <i>et al.</i> 2017 (68)</p> <p>Indeterminate (inadequate statistical power to detect low frequency variant):</p>	High

			Amstutz, <i>et al.</i> 2013 (37)	
Clinical	SJS/TEN	Cases of patients with oxcarbazepine-induced SJS/TEN and <i>HLA-B*15:02</i> genotype.	Chen, <i>et al.</i> 2009 (73) Hung, <i>et al.</i> 2010 (74) Sun, <i>et al.</i> 2014 (45)	Moderate
Clinical	DRESS	Case of a patient with oxcarbazepine-induced DRESS and <i>HLA-B*15:02</i> genotype.	Shankarkumar, <i>et al.</i> 2009 (75)	Weak
Clinical	MPE	Cases of patients with oxcarbazepine-induced MPE and <i>HLA-B*15:02</i> genotype.	Wang, <i>et al.</i> 2012 (64) Wang, <i>et al.</i> 2014 (72)	Weak

DRESS: drug reaction with eosinophilia and systemic symptoms; MPE: maculopapular exanthema; SJS: Stevens-Johnson syndrome;

TEN: toxic epidermal necrolysis

^aRating scheme described in the **Supplemental Material**.

TABLE S2. EVIDENCE LINKING *HLA-A*31:01* GENOTYPE WITH CARBAMAZEPINE- AND OXCARBAZEPINE-INDUCED CUTANEOUS ADVERSE REACTIONS

Type of Experimental Model	Clinical Phenotype	Major Findings	References	Level of Evidence ³
Carbamazepine and <i>HLA-A*31:01</i>				
In vitro	N/A	HLA-A*31:01 restricted the activation of carbamazepine-specific CD8(+) T-cells that were derived from a patient with <i>HLA-A*31:01</i> genotype who presented with a generalized maculopapular exanthema with eosinophilia and lymphocytosis 6 days after starting carbamazepine.	Lichtenfels, <i>et al.</i> 2014 (76)	Weak
Clinical	DRESS	Significant association between <i>HLA-A*31:01</i> genotype and patients with carbamazepine-induced DRESS compared to carbamazepine-tolerant patients and/or healthy controls.	Supports statement: Hung, <i>et al.</i> 2006 (25) Kashiwagi, <i>et al.</i> 2008 (54) Kim, <i>et al.</i> 2011 (9) McCormack, <i>et al.</i> 2011 (77) Ozeki, <i>et al.</i> 2011 (78) Niihara, <i>et al.</i> 2012 (58)	High

			<p>Amstutz, <i>et al.</i> 2013 (37)</p> <p>Genin, <i>et al.</i> 2014 (46)</p> <p>Hsiao, <i>et al.</i> 2014 (47)</p> <p>Indeterminate (inadequate statistical power to detect low frequency variant):</p> <p>Shirzadi, <i>et al.</i> 2015 (79)</p>	
Clinical	MPE	Significant association between <i>HLA-A*31:01</i> genotype and patients with carbamazepine-induced MPE compared to carbamazepine-tolerant patients and/or healthy controls.	<p>Supports statement:</p> <p>Hung, <i>et al.</i> 2006 (25)</p> <p>Kashiwagi, <i>et al.</i> 2008 (54)</p> <p>Hsiao, <i>et al.</i> 2014 (47)</p> <p>McCormack, <i>et al.</i> 2011 (77)</p> <p>Ozeki, <i>et al.</i> 2011 (78)</p> <p>Niihara, <i>et al.</i> 2012 (58)</p>	Moderate

			<p>Amstutz, <i>et al.</i> 2013 (37)</p> <p>Fricke-Galindo, <i>et al.</i> 2014 (80)</p> <p>Indeterminate (inadequate statistical power to detect low frequency variant):</p> <p>Li, <i>et al.</i> 2013 (60)</p> <p>Song, <i>et al.</i> 2014 (81)</p> <p>Shirzadi, <i>et al.</i> 2015 (79)</p>	
Clinical	SJS/TEN	Significant association between <i>HLA-A*31:01</i> genotype and patients with carbamazepine-induced SJS/TEN compared to carbamazepine-tolerant patients.	<p>Supports statement:</p> <p>Ozeki, <i>et al.</i> 2011 (78)</p> <p>McCormack, <i>et al.</i> 2011 (77)</p> <p>Genin, <i>et al.</i> 2014 (46)</p> <p>Indeterminate (inadequate statistical power to</p>	High

			<p>detect low frequency variant): Hung, <i>et al.</i> 2006 (25) Kim, <i>et al.</i> 2011 (9) Niihara, <i>et al.</i> 2012 (58) Shi, <i>et al.</i> 2012 (35) Amstutz, <i>et al.</i> 2013 (37) Genin, <i>et al.</i> 2014 (46) Hsiao, <i>et al.</i> 2014 (47) Park, <i>et al.</i> 2016 (59)</p>	
Clinical	DRESS	Cases of patients with carbamazepine-induced DRESS and <i>HLA-A*31:01</i> genotype.	Mizumoto, <i>et al.</i> 2012 (82) Anjum, <i>et al.</i> 2014 (83) Segert, <i>et al.</i> 2016 (84)	Weak
Oxcarbazepine and <i>HLA-A*31:01</i>				
Clinical	DRESS/MPE	No significant association between <i>HLA-A*31:01</i> genotype and patients with oxcarbazepine-induced non-SJS/TEN cutaneous	Supports statement: Chen, <i>et al.</i> 2017 (68)	Moderate

		adverse drug reaction compared to oxcarbazepine-tolerant patients or healthy controls.	Indeterminate (inadequate statistical power to detect low frequency variant): Amstutz, <i>et al.</i> 2013 (37)	
Clinical	SJS/TEN	No significant association between <i>HLA-A*31:01</i> genotype and patients with oxcarbazepine-induced SJS/TEN compared to oxcarbazepine-tolerant patients or healthy controls.	Supports statement: Chen, <i>et al.</i> 2017 (68) Indeterminate (inadequate statistical power to detect low frequency variant): Amstutz, <i>et al.</i> 2013 (37)	High

DRESS = drug reaction with eosinophilia and systemic symptoms; MPE = maculopapular exanthema; SJS = Stevens-Johnson syndrome; TEN = toxic epidermal necrolysis

^aRating scheme described in the **Supplemental Material**

HLA-B Reference ATGCTGGTCATGGCGCCCCGAACCGTCCTCCTGCTGCTCTCGGCGGCCCTGGCCCTGACCGAGACCTGGGCCGGCTCCCACTCCATGAGGTATTTCTACA
HLA-B*1502 ATGC**GGGTCA**CGGCGCCCCGAACCGTCCTCCTGCTGCTCTCGG**GAG**CCCTGGCCCTGACCGAGACCTGGGCCGGCTCCCACTCCATGAGGTATTTCTACA

HLA-B Reference CCTCCGTGTCCCGGCCCGGCCGCGGGGAGCCCCGCTTCATCTCAGTGGGCTACGTGGACGACACCCAGTTCGTGAGGTTTCGACAGCGACGCCGCGAGTCC
HLA-B*1502 CC**GCC**ATGTCCCGGCCCGGCCGCGGGGAGCCCCGCTTCATC**G**CAGTGGGCTACGTGGACGACACCCAGTTCGTGAGGTTTCGACAGCGACGCCGCGAGTCC

HLA-B Reference GAGAGAGGAGCCGCGGGCGCCGTGGATAGAGCAGGAGGGGCGGAGTATTGGGACCGGAACACACAGATCTACAAGGCCAGGCACAGACTGACCGAGAG
HLA-B*1502 GAG**GAT**GG**CGCC**CCGGGCGCC**A**TGGATAGAGCAGGAGGGGCGGAGTATTGGGACCGGAACACACAGATCT**CCAAGACCACA**CACAGACT**T**ACCGAGAG

HLA-B Reference AGCCTGCGGAACCTGCGCGGCTACTACAACCAGAGCGAGGCCGGGTCTCACACCCTCCAGAGCATGTACGGCTGCGACGTGGGGCCGGACGGGGCGCCTCC
HLA-B*1502 AGCCTGCGGAACCTGCGCGGCTACTACAACCAGAGCGAGGCCGGGTCTCACAT**TC**ATCCAGAG**G**ATGTAT**T**GGCTGCGACGTGGGGCCGGACGGGGCGCCTCC

HLA-B Reference TCCGCGGGCATGACCAGTACGCCTACGACGGCAAGGATTACATCGCCCTGAACGAGGACCTGCGCTCCTGGACCGCCGCGGACACGGCGGCTCAGATCAC
HLA-B*1502 TCCGCGGG**T**ATGACCAGT**CC**GCCTACGACGGCAAGGATTACATCGCCCTGAACGAGGACCT**G**AGCTCCTGGACCGC**GG**CGGACACGGCGGCTCAGATCAC

HLA-B Reference CCAGCGCAAGTGGGAGGCGGCCCGTGAGGCGGAGCAGCGGAGAGCCTACCTGGAGGGCGAGTGCCTGGAGTGGCTCCGCAGATACCTGGAGAACGGGAAG
HLA-B*1502 CCAGCGCAAGTGGGAGGCGGCCCGTGAGGCGGAGCAGC**T**GAGAGCCTACCTGGAGGGC**CT**GTGCGTGGAGTGGCTCCGCAGATACCTGGAGAACGGGAAG

HLA-B Reference GACAAGCTGGAGCGCGCTGACCCCCCAAAGACACACGTGACCCACCACCCCATCTCTGACCATGAGGCCACCCTGAGGTGCTGGGCCCTGGGTTTCTACC
HLA-B*1502 GAG**AC**GCTG**C**AGCGCGC**G**GACCCCCCAAAGACACAT**T**GTGACCCACCACCCCATCTCTGACCATGAGGCCACCCTGAGGTGCTGGGCCCTGGG**C**TTCTACC

HLA-B Reference CTGCGGAGATCACACTGACCTGGCAGCGGGATGGCGAGGACCAAACCTCAGGACACTGAGCTTGTGGAGACCAGACCAGCAGGAGATAGAACCTTCCAGAA
HLA-B*1502 CTGCGGAGATCACACTGACCTGGCAGCGGGATGGCGAGGACCAAACCTCAGGACAC**C**GAGCTTGTGGAGACCAGACCAGCAGGAGATAGAACCTTCCAGAA

HLA-B Reference GTGGGCAGCTGTGGTGGTGCCTTCTGGAGAAGAGCAGAGATACACATGCCATGTACAGCATGAGGGGCTGCCGAAGCCCCTCACCTGAGATGGGAGCCG
HLA-B*1502 GTGGGCAGCTGTGGTGGTGCCTTCTGGAGAAGAGCAGAGATACACATGCCATGTACAGCATGAGGGGCTGCCGAAGCCCCTCACCTGAGATGGGAGCC**A**

HLA-B Reference TCTTCCAGTCCACCGTCCCCATCGTGGGCATTGTTGCTGGCCTGGCTGTCTTAGCAGTTGTGGTCATCGGAGCTGTGGTCGCTGCTGTGATGTGTAGGA
HLA-B*1502 TCTTCCAGTCCAC**C**ATCCCCATCGTGGGCATTGTTGCTGGCCTGGCTGTCTTAGCAGTTGTGGTCATCGGAGCTGTGGTCGCT**A**CTGTGATGTGTAGGA

HLA-B Reference GGAAGAGTTCAGGTGGAAAAGGAGGGAGCTACTCTCAGGCTGCGTGCAGCGACAGTGCCCAGGGCTCTGATGTGTCTCTCACAGCTTGA
HLA-B*1502 GGAAGAG**C**TAGGTGGAAAAGGAGGGAGCTACTCTCAGGCTGCGTGCAGCGACAGTGCCCAGGGCTCTGATGTGTCTCTCACAGCTTGA

Figure S1. Nucleotide coding sequence alignment of *HLA-B*15:02* and the reference sequence. Nucleotide differences between the two sequences are highlighted in blue. This alignment was generated using the IMGT/HLA Database's alignment tool (www.ebi.ac.uk/imgt/hla/align.html) and visualized in Jalview (85).

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HLA-B Reference  MLVMAPRTVLLLLSAALALTETWAGSHSMRYFYTSVSRPGRGEPRFISVG
HLA-B*1502       MRVTAPRTVLLLLSGALALALTETWAGSHSMRYFYTAMSRPGRGEPFIAVG

HLA-B Reference  YVDDTQFVRFDSDAASPREEPRAPWIEQEGPEYWRNTQIYKAQAQTDRE
HLA-B*1502       YVDDTQFVRFDSDAASPMAPRAPWIEQEGPEYWRNTQISKTNTQTYRE

HLA-B Reference  SLRNLRGYYNQSEAGSHTLQSMYGC DVGPDGRLLRGHDQYAYDGKDYIAL
HLA-B*1502       SLRNLRGYYNQSEAGSHIIQRMYGC DVGPDGRLLRGYDQSAYDGKDYIAL

HLA-B Reference  NEDLRSWTAADTAAQITQRKWEAAREAEQRRAYLEGECEVWLRRYLENGK
HLA-B*1502       NEDLSSWTAADTAAQITQRKWEAAREAEQLRAYLEGLCEVWLRRYLENGK

HLA-B Reference  DKLERADPPKTHVTHHPISDHEATLRCWALGFYPAEITLTWQRDGEDQTQ
HLA-B*1502       ETLQRADPPKTHVTHHPISDHEATLRCWALGFYPAEITLTWQRDGEDQTQ

HLA-B Reference  DTELVETRPAGDRTFQKWA AVVVP SGEEQRYTCHVQHEGLPKPLTLRWEP
HLA-B*1502       DTELVETRPAGDRTFQKWA AVVVP SGEEQRYTCHVQHEGLPKPLTLRWEP

HLA-B Reference  SSQSTVPIVGIVAGLAVLAVVVIGAVVA VMCRKSSGGKGGSYSQAACS
HLA-B*1502       SSQSTIPIVGIVAGLAVLAVVVIGAVVATVMCRKSSGGKGGSYSQAASS

HLA-B Reference  DSAQGS DVSLTA
HLA-B*1502       DSAQGS DVSLTA

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Figure S2. Amino acid sequence alignment of *HLA-B*15:02* and the reference sequence. Amino acid differences between the two sequences are highlighted in blue. This alignment was generated using the IMGT/HLA Database's alignment tool (www.ebi.ac.uk/imgt/hla/align.html) and visualized in Jalview (85).

cDNA	10	20	30	40	50	60	70	80	90	100
A*01:01:01:01	ATGGCCGTCA	TGGCGCCCG	AACCCCTCCTC	CTGCTACTCT	CGGGGGCCCT	GGCCCTGACC	CAGACCTGGG	CGGGCTCCCA	CTCCATGAGG	TATTTCTTCA
A*31:01:02:01	ATGGCCGTCA	TGGCGCCCG	AACCCCTCCTC	CTGCTACTCT	TGGGGGCCCT	GGCCCTGACC	CAGACCTGGG	CGGGCTCCCA	CTCCATGAGG	TATTTCA ACCA
cDNA	110	120	130	140	150	160	170	180	190	200
A*01:01:01:01	CATCCGTGTC	CCGGCCCGG	CGCGGGGAGC	CCCGTTTCAT	CGCCGTGGGC	TACGTGGACG	ACACGCAGTT	CGTGC GGTTT	GACAGCGACG	CCGCAGCCA
A*31:01:02:01	CATCCGTGTC	CCGGCCCGG	CGCGGGGAGC	CCCGTTTCAT	CGCCGTGGGC	TACGTGGACG	ACACGCAGTT	CGTGC GGTTT	GACAGCGACG	CCGCAGCCA
cDNA	210	220	230	240	250	260	270	280	290	300
A*01:01:01:01	GAAGATGGAG	CCGCGGGCGC	CGTGGATAGA	GCAGGAGGG	CCGGAGTATT	GGGACCAGGA	GACACGGAAT	ATGAAGGCC	ACTCACAGAC	TGACCGAGCG
A*31:01:02:01	GAGGATGGAG	CCGCGGGCGC	CGTGGATAGA	GCAGGAGAG	CC T GAGTATT	GGGACCAGGA	GACACGGAAT	G TGAAGGCC	ACTCACAGAT	TGACCGAG TG
cDNA	310	320	330	340	350	360	370	380	390	400
A*01:01:01:01	AACCTGGGGA	CCCTGCGCG	CTACTACAAC	CAGAGCGAGG	ACGGTTCTCA	CACCATCCAG	AT A ATGTATG	GCTGCGACGT	GGGG C CGGAC	GGGCCTTCC
A*31:01:02:01	GACCTGGGGA	CCCTGCGCG	CTACTACAAC	CAGAGCGAGG	CCGGTTCTCA	CACCATCCAG	AT G ATGTATG	GCTGCGACGT	GGGG T CGGAC	GGGCCTTCC
cDNA	410	420	430	440	450	460	470	480	490	500
A*01:01:01:01	TCCGCGGGTA	CCGGCAGGAC	GCCTACGACG	GCAAGGATTA	CATCGCCCTG	AACGAGGACC	TGCGCTCTTG	GACCGCGGCG	GACATGGC A G	CTCAGATCAC
A*31:01:02:01	TCCGCGGGTA	CC A GAGGAC	GCCTACGACG	GCAAGGATTA	CATCGC C TG	AACGAGGACC	TGCGCTCTTG	GACCGCGGCG	GACATGGC G	CTCAGATCAC
cDNA	510	520	530	540	550	560	570	580	590	600
A*01:01:01:01	CAAGCGCAAG	TGGGAGGCGG	TCCATGCGGC	GGAGCAGCGG	AGAGTCTACC	TGGAGGGCG	GTGCGTGGAC	GGGCTCCGCA	GATACCTGGA	GAACGGGAAG
A*31:01:02:01	CCAGCGCAAG	TGGGAGGCGG	CC C G T GCGG	GGAGCAG T TG	AGAG C CTACC	TGGAGGG C A	GTGCGTGGAG	T GGCTCCGCA	GATACCTGGA	GAACGGGAAG
cDNA	610	620	630	640	650	660	670	680	690	700
A*01:01:01:01	GAGACGCTGC	AGCGCACGGA	CCCCCCAAG	ACACATATGA	CCACCACCC	CA T CTCTGAC	CATGAGGCCA	CCCTGAGGTG	CTGGGCCCTG	GGCTTCTACC
A*31:01:02:01	GAGACGCTGC	AGCGCACGGA	CCCCCCAAG	AC G CATATGA	CTCACCAC G C	T GTCTCTGAC	CATGAGGCCA	CCCTGAGGTG	CTGGGCCCTG	AGCTTCTACC
cDNA	710	720	730	740	750	760	770	780	790	800
A*01:01:01:01	CTGCGGAGAT	CACACTGACC	TGGCAGCGGG	ATGGGGAGGA	CCAGACCCAG	GACACGGAGC	TCGTGGAGAC	CAGGCCTGCA	GGGGATGGAA	CCTTCCAGAA
A*31:01:02:01	CTGCGGAGAT	CACACTGACC	TGGCAGCGGG	ATGGGGAGGA	CCAGACCCAG	GACACGGAGC	TCGTGGAGAC	CAGGCCTGCA	GGGGATGGAA	CCTTCCAGAA
cDNA	810	820	830	840	850	860	870	880	890	900
A*01:01:01:01	GTGGGCGGCT	GTGGTGGTGC	CTTCTGGAG	GGAGCAGAGA	TACACCTGCC	ATGTGCAGCA	TGAGGGTCTG	CCCAAGCCCC	TCACCTGAG	ATGGGAGCTG
A*31:01:02:01	GTGGGCG T CT	GTGGTGGTGC	CTTCTGG A CA	GGAGCAGAGA	TACACCTGCC	ATGTGCAGCA	TGAGGGTCT C	CCCAAGCCCC	TCACCTGAG	ATGGGAG CC G
cDNA	910	920	930	940	950	960	970	980	990	1000
A*01:01:01:01	TCTTCCCAGC	CCACCATCCC	CATCGTGGGC	ATCATTGCTG	GCCTGGTTCT	CC T TGGAGCT	GTG A TCACTG	GAGCTGTGGT	CGCTGC C GTG	ATGTGGAGGA
A*31:01:02:01	TCTTCCCAGC	CCACCATCCC	CATCGTGGGC	ATCATTGCTG	GCCT A GTTTCT	CT T TGGAGCT	GTG T T C GCTG	GAGCTGTGGT	CGCTGC T GTG	AGGTGGAGGA
cDNA	1010	1020	1030	1040	1050	1060	1070	1080	1090	
A*01:01:01:01	GGAAGAGCTC	AGATAGAAAA	GGAGGGAG T T	ACACTCAGGC	TGCAAGCAGT	GACAGTGCC	AGGGCTCTGA	T G TGTCTCTC	ACAGCTTGTA	AAGTGTA
A*31:01:02:01	GGAAGAGCTC	AGATAGAAAA	GGAGGGAG C T	AC T CTCAGGC	TGCAAGCAGT	GACAGTGCC	AGGGCTCTGA	T A T G T CTCTC	ACAGCTTGTA	AAGTGTA

Figure S3. Nucleotide coding sequence alignment of *HLA-A*31:01* and the reference sequence (*HLA-A*01:01*). Nucleotide differences between the two sequences are highlighted in red. This alignment was generated using the IMGT/HLA Database's alignment tool (www.ebi.ac.uk/imgt/hla/align.html) and highlighted manually.

AA Pos.	-21	-11	-1	10	20	30	40	50	60	70	
A*01:01:01:01	MAVM	APRTL	LLLLLS	GALALTQTWA	GSHSMRYF FT	SVSRPGRGEP	RFIAVGYVDD	TQFVRFDSDA	ASQ K MEPRAP	WIEQE G PEYW	DQETRN M KAH
A*31:01:02:01	MAVM	APRTL	LLLLLL	GALALTQTWA	GSHSMRYF TT	SVSRPGRGEP	RFIAVGYVDD	TQFVRFDSDA	ASQ R MEPRAP	WIEQE R PEYW	DQETRN V KAH
AA Pos.	80	90	100	110	120	130	140	150	160	170	
A*01:01:01:01	SQ T DRANLGT	LRGYNQSE D	GSHTIQ I MYG	CDVGP D GRFL	RGY R QDAYDG	KDYIALNEDL	RSWTAADMAA	QIT K RKWEAV	H AAEQ R RVYL	EG R CVDGLRR	
A*31:01:02:01	SQ I DR V DLGT	LRGYNQSE A	GSHTIQ M MYG	CDVGS D GRFL	RGY Q QDAYDG	KDYIALNEDL	RSWTAADMAA	QIT Q RKWEA A	R VAEQ L RAYL	EG T CV E WLR	
AA Pos.	180	190	200	210	220	230	240	250	260	270	
A*01:01:01:01	YLENGKETLQ	RTDPPKTHMT	HH P ISDHEAT	LRCWAL G FYP	AEITLTWQRD	GEDQTQDTEL	VETRPAGDGT	FQKWA A VVVP	SG E EQRYTCH	VQHEGLPKPL	
A*31:01:02:01	YLENGKETLQ	RTDPPKTHMT	HH A VSDHEAT	LRCWAL S FYP	AEITLTWQRD	GEDQTQDTEL	VETRPAGDGT	FQKWA S VVVP	SG Q EQRYTCH	VQHEGLPKPL	
AA Pos.	280	290	300	310	320	330	340				
A*01:01:01:01	TLRWE L SSQP	TIPVGGIAG	LVL L GAV I TG	AVVA A V M WRR	KSSDRKGGSY	T QAASSDSAQ	GSD V SLTACK	V			
A*31:01:02:01	TLRWE P SSQP	TIPVGGIAG	LVL F GAV F AG	AVVA A V R WRR	KSSDRKGGSY	S QAASSDSAQ	GSD M SLTACK	V			

Figure S4. Amino acid sequence alignment of HLA-A*31:01 and the reference sequence (HLA-A*01:01). Amino acid differences between the two sequences are highlighted in red. This alignment was generated using the IMGT/HLA Database's alignment tool (www.ebi.ac.uk/imgt/hla/align.html) and highlighted manually.

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